

UniversitätsSpital Zürich
Klinik für Nephrologie
Direktor: Prof. Dr. med. R. P. Wüthrich

Arbeit unter Leitung von Dr. med. D. Franzen und Dr. med. M. Bonani

Chronic norovirus infection is a risk factor for secondary lactose intolerance in renal transplant recipients. A case-control study

INAUGURAL-DISSERTATION

zur Erlangung der Doktorwürde der Humanmedizin
der Medizinischen Fakultät
der Universität Zürich

vorgelegt von
Rajha Messias Fabrizio Pereira
von Zürich

Genehmigt auf Antrag von Prof. Dr. med. R. P. Wüthrich
Zürich 2016

Publikationshinweis

Chronic norovirus infection is a risk factor for secondary lactose intolerance in renal transplant recipients. A case-control study.

Publiziert am: 01. August 2016

Journal: Transplantation
http://journals.lww.com/transplantjournal/Abstract/onlinefirst/Chronic_norovirus_infection_as_a_risk_factor_for.97297.aspx

Pubmed
<http://www.ncbi.nlm.nih.gov/pubmed/27482964>

Publikation

**Chronic norovirus infection as a risk factor for secondary lactose
maldigestion in renal transplant recipients: a prospective parallel
cohort pilot study**

Marco Bonani, MD^{1,*}; Rahja M. Pereira, MD^{1,*}; Benjamin Misselwitz, MD²;

Thomas Fehr, MD^{1,3}; Rudolf P. Wüthrich, MD¹; Daniel Franzen, MD⁴

*The first 2 authors contributed equally to this article

¹ Department of Nephrology, ² Department of Gastroenterology and Hepatology, University Hospital Zürich, Zürich, Switzerland; ³ Department of Internal Medicine, Cantonal Hospital Graubünden, Chur, Switzerland; ⁴ Department of Pulmonology, University Hospital Zürich, Zürich, Switzerland

Trial registration: This study is registered at ClinicalTrials.gov (identifier: NCT01840891)

Corresponding author:

Dr Daniel Franzen

Department of Pulmonology, University Hospital Zürich

Rämistrasse 100, 8091 Zürich

Switzerland

Phone: + 41 44 255 11 11, Fax: + 41 44 255 44 51

Email: daniel.franzen@usz.ch

AUTHORSHIP PAGE

Authors' contributions

Study design (TF, RPW, DF), data collection (RP, MB), data analysis (MB, BM, DF), drafting of the manuscript (MB, BM, TF, RPW, DF). Approval of the final version of the manuscript (all authors).

Disclosures

The authors declare no conflicts of interest

Funding

There is no funding source of this study

ABBREVIATIONS PAGE

CMV, cytomegalovirus

IBS, irritable bowel syndrome

IQR, interquartile range

LHBT, lactose hydrogen breath test

LTT, lactose tolerance test

LCT, lactase

LI, lactose intolerance

LM, lactose maldigestion

PCR, polymerase chain reaction

RTR, renal transplant recipient

PCR, polymerase chain reaction

SIBO, small intestine bacterial overgrowth

ABSTRACT

Background: Chronic norovirus infection is an emerging challenge in the immunocompromised host, in whom it may be asymptomatic or present as chronic diarrhea. The mechanisms of diarrhea in chronic norovirus infection are not well understood, but in analogy to *Gardia lamblia* and rotavirus infections, secondary lactose maldigestion (LM) might be implicated.

Methods: Adult renal transplant recipients (RTRs) who had symptomatic chronic norovirus infection with diarrhea were asked to participate in this prospective parallel cohort study. RTRs with otherwise unexplainable chronic diarrhea but absent infection served as control group. In both groups, a lactose hydrogen breath test (LHBT) and a lactose tolerance test (LTT) were performed after exclusion of primary LM by a negative lactase gene test.

Results: Of approximately 800 patients in the cohort of RTRs at our institution, 15 subjects were included in the present study. Of these, 7 had chronic symptomatic norovirus infection with diarrhea (noro group) and 8 had diarrhea in the absence of norovirus (control group). LHBT and LTT were positive in all 7 patients (100%) in the noro group, whereas only 1 of 8 patients (12.5%) in the control group had a positive test. Thus, secondary LM was highly prevalent in the noro compared to the control group with an odds ratio of 75.0 (95% CI 2.6, 2153, $p=0.01$).

Conclusions: This is the first report showing a positive association of chronic norovirus infection and secondary LM. Further studies with larger patient numbers and longer follow-up are needed to test a causative relationship between both entities.

Key Words: Chronic norovirus infection, chronic diarrhea, renal transplant recipients, lactose maldigestion

Introduction

Norovirus, a single-stranded RNA virus of the Caliciviridae family, is a human enteric pathogen that is 1 of the leading causes of acute gastroenteritis, presenting as self-limited disease of short duration in immunocompetent subjects.¹⁻³ However, chronic norovirus infection is an emerging challenge in the immunocompromised host such as leukemia patients or solid organ transplant recipients, in whom the virus may persist and present as chronic diarrhea and diffuse abdominal discomfort, and may even be associated with kidney transplant dysfunction.⁴⁻⁸ Norovirus accounts for 17-26% of severe posttransplant diarrhea in renal transplant recipients.^{5,6,9} Norovirus related diarrhea is associated with the greatest weight loss compared to other causes of diarrhea.^{5,9} Histologically, signs of chronic intestinal inflammation are present.^{4,5} Until now, the mechanisms of diarrhea in case of chronic norovirus infection are not well understood, and treatment options are limited.

Lactose is a disaccharide and a frequent constituent of a typical Western-type diet. Lactose maldigestion (LM) refers to inefficient cleavage of lactose in the small intestine, resulting in lactose malabsorption and fermentation of lactose by the colonic microbiota. In contrast, lactose intolerance (LI) is defined as the development of symptoms after lactose challenge in individuals with LM.¹⁰ LM is a frequent condition, affecting more than 50% of all individuals worldwide and should be regarded a variant of human intestinal physiology.¹¹ Primary LM is typically associated with the CC polymorphism of the -13910 locus of the lactase (LCT) gene.¹² In contrast, secondary LM can develop in many intestinal inflammatory conditions; however, which specific conditions will lead to LM as well as mechanistic aspects, have not been sufficiently clarified.

In a prospective study in children with acute gastroenteritis, a significant proportion was found to have LM, which was most commonly associated with rotavirus infection.¹³ Secondary LM has also been reported in patients with *Giardia lamblia* infections, and the latter were shown to alter the cellular glycocalyx resulting in alterations of brush border disaccharidase enzymes.^{14,15} In line with these findings we suspect a similar mechanism in symptomatic patients with chronic norovirus infections. The objective of this study was therefore to determine the prevalence of secondary LM in patients with chronic norovirus infection.

Materials and Methods

Subjects

Between July 2013 and March 2015 all adult renal transplant recipients (RTRs) at the University Hospital Zürich who had symptomatic chronic norovirus infection with diarrhea were asked to participate in this prospective parallel cohort study. According to the WHO-approved definition of diarrhea we chose the cut-off of 3 or more bowel movements per day for more than 4 weeks as indicative of chronic diarrhea. Chronic norovirus infection was proven by positive polymerase chain reaction (PCR) analysis of recent stool samples, whereas chronic virus shedding was defined as more than 2 PCR positive samples at an interval of at least 1 month. Concomitant viral (ie cytomegalovirus), bacterial (ie *Salmonella* spp., *Campylobacter* spp., *Shigella* spp., and *C.difficile*) and parasitic (*Giardia lamblia*, *Microspora* spp., and *Cryptospora* spp.) intestinal infections were excluded by negative stool PCR analyses, stool cultures, and direct microscopic stool examinations, respectively. Furthermore, CMV viremia was excluded by PCR technique. Main exclusion criterion for the present study was a concomitant intestinal infection (other than norovirus), and primary LM which was previously excluded by absence of

the CC genotype of the DNA variant -13910 T/C upstream in the LCT gene. Subjects with a proven galactosemia or those requiring a low galactose diet were also excluded. RTRs with otherwise unexplainable chronic diarrhea but absent norovirus or another intestinal infection, and negative LCT gene test served as control group. In both groups, a lactose hydrogen breath test (LHBT) and a lactose tolerance test (LTT) were performed in all eligible RTRs (Figure 1). LM was diagnosed with a positive LHBT and/or a positive LTT.

Written informed consent was obtained from all patients included in the study. The study was approved by the local Ethics committee (KEK-ZH 2012-0473) and is registered at ClinicalTrials.gov (identifier: NCT01840891).

Lactose H₂ breath test (LHBT)

The LHBT was performed according to the Rome consensus conference.¹⁶ After an overnight fast of at least 12 hours, a basal breath sample was collected. No individual showed a baseline hydrogen (H₂) level above 20 ppm (not shown). RTRs were allowed to drink water and follow their usual medication regimen during the entire examination. After collecting the baseline sample, RTRs were given 25 g of lactose dissolved in 250 ml of water to drink. Orange flavored lactose powder (or milk powder) was provided with the AlveoSampler™ Lactose Kit (Quintron Instrument Co., Milwaukee, WI, USA). Samples of end expiratory breath were then collected at 30, 60, 90 and 120 minutes after the oral lactose load to measure the concentration of H₂, which was considered significantly increased and indicative of LM when exceeding 20 ppm.^{16,17} During the test, RTRs were allowed to engage in normal activities, but were kept fasting except for water consumption which was permitted throughout the examination. The test was performed in a well-ventilated room free of fresh painted walls or objects and with no

evidence of any organic solvents or cigarette smoke. The breath samples were collected in specially constructed bags, which are provided along with the instrument. Exhaled breath H_2 was measured on a Model 12i Microlyser (Quintron Instrument Co., Milwaukee, WI, USA). The number of loose bowel motions and flatulence during the test were also documented.

Lactose tolerance test (LTT)

Following the above mentioned oral administration of 25 g lactose, capillary blood glucose levels were measured at 0, 60, and 120 minutes, by using a glucometer (Ascensia Contour, Bayer AG, Leverkusen, Germany). An increase of blood glucose by less than 1.1 mmol/l in conjunction with the development of abdominal symptoms was defined diagnostic for LI.¹⁷

Statistical analysis

Baseline data are reported as median (interquartile range, IQR), or numbers (percentages) as appropriate. Differences in baseline characteristics and between the 2 study groups were estimated using the Mann-Whitney U test for continuous variables and the χ^2 test for categorical variables. Values of exhaled breath hydrogen concentrations and blood glucose are presented as median (interquartile range, IQR). Differences of these values between different points in time were calculated using the paired sample Wilcoxon signed rank test. *P*-values of all outcomes were 2-sided; values less than 0.05 were considered to indicate statistical significance. All statistical analyses were performed using IBM SPSS Statistics for Windows, version 22 (IBM Corporation, Armonk, NY).

Results

Of approximately 800 patients in the cohort of adult RTRs at the University Hospital Zürich, 22 individuals were identified with chronic diarrhea, 15 of which could be included in the present study (Figure 1). Four individuals were excluded due to primary LM in line with an expected frequency of primary LM in Switzerland of 20-40%¹¹. Seven of these 15 individuals had chronic symptomatic norovirus infection with diarrhea with the genotype G2.4 (noro group), and 8 patients had diarrhea in the absence of norovirus infection and served as control group. Baseline characteristics of the subjects in both groups are shown in Table 1, and laboratory data are summarized in Table 2. Cytomegalovirus high risk constellation (CMV donor/recipient serostatus D+/R-) was significantly more prevalent in the control group ($p=0.013$). However, the onset of diarrhea before study inclusion was significantly earlier in the noro group ($p=0.038$). Other variables, such as age, sex, mode and dose of immunosuppression, prevalence of diabetes mellitus, and laboratory values were comparable in both groups. At the moment of the testing, no patient was treated with antibiotics, and CMV PCR was negative in all patients. As part of the clinical routine diagnostic in transplanted patients with chronic diarrhea, 6 patients (85.7%) in the norovirus group had a colonoscopy. In all patients, the histology showed chronic inflammatory changes. CMV-colitis was specifically excluded with histology and immunohistochemistry of biopsy specimens. The final diagnosis of chronic diarrhea in those patients without evidence of norovirus infection was mycophenolate-associated colitis in 3 and unknown etiology or diabetes mellitus in 5 cases.

In the noro group, all patients had a positive LHBT. In the control group, only 1 patient (12.5%) had a positive test (Table 3). Accordingly, the increase of the median exhaled H₂ content

between baseline (7.0 (2.0, 11.0) ppm) and 120 minutes after lactose ingestion (35.5 (10.0, 66.2) ppm) in the noro group was significant ($p=0.043$) (Figure 2). By contrast, in the control group H_2 values only a minor, nonsignificant increase between baseline (4.0 (1.2, 6.0) ppm) and after lactose exposure was observed (9.0 (4.0, 16.2) ppm) ($p=0.063$) and all values remained below the threshold of 20ppm.

Analogously, LTT was positive in all patients in the noro group, whereas only the above mentioned patient in the control group had a positive test (Table 3). However, in both groups the increase of blood glucose between baseline and 60 min after lactose ingestion was significant (Figure 3), although the difference was more pronounced in the control group (baseline 5.3 (5.1, 6.6) mmol/l; after 60 min 7.9 (5.9, 8.5) mmol/l) ($p=0.017$) compared to the noro group (baseline 5.4 (4.8, 5.5) mmol/l; after 60 min 6.4 (5.7, 7.0) mmol/l) ($p=0.046$).

In all but 1 patient of the noro group, there were abdominal symptoms after lactose ingestion (diarrhea, $n=2$; bloating, $n=3$; combination of diarrhea and bloating, $n=1$). In the control group, no patient reported abdominal symptoms (Table 3). The patient in the control group with positive LHBT and LTT denied any symptoms after lactose ingestion.

Based on both tests, secondary LM was highly prevalent in the noro group compared to the control group with an odds ratio of 75.0 (95% CI 2.6, 2153), $p=0.01$. Likewise, for secondary LI (defined as LM with symptoms), the odds ratio was 73.7 (95% CI 2.6, 2120), $p=0.01$ (Table 3).

Discussion

Chronic norovirus infection is an emerging challenge in the immunocompromised host, in whom it may present as chronic diarrhea. The aim of the present study was to investigate

whether secondary LM can contribute to diarrhea in patients with chronic norovirus shedding. This is the first report showing a positive association of chronic norovirus infection in RTRs and secondary LM, suggesting a causative relationship between both entities. In addition, LI was highly prevalent, and diarrhea lasted substantially longer in RTRs with symptomatic chronic norovirus infection. Thus, secondary LM due to chronic norovirus infection could possibly be another cause of chronic diarrhea beside drug-induced diarrhea (eg mycophenolate) in immunosuppressed patients. Schorn et al found in their case series of RTRs with chronic norovirus infection, that the intensity of immunosuppression correlated with diarrheal symptoms but not with viral shedding.⁶ Thus, immunosuppression dosage is maybe the most important risk factor for chronic norovirus infection. Therefore, we generally follow a stepwise approach with first reducing the dosage of mycophenolate because of the possibility of a coincident mycophenolate toxicity contributing to the chronic diarrhea, followed by reduction of the calcineurin-inhibitor dosage and attempt to taper/stop prednisone therapy. In our study, immunosuppressant dosage was similar in both groups in our study. However, lymphocyte counts were significantly lower in the noro group.

In general, endoscopy was performed for ongoing chronic diarrhea to rule out other conditions. However, colonoscopy was not an inclusion criterion in this study, although most of the patients had the procedure performed prior to study inclusion. Our standard procedure in RTR's with chronic diarrhea is CMV-PCR in blood, stool cultures for bacteria including *Clostridium difficile* and *Clostridium* toxin detection, PCR analysis of stool specimens for norovirus, and microscopy for parasites such as *Microsporidium* and *Cryptosporidium*. If the tests are negative, and the diarrhea persists, we first reduce the mycophenolate doses or change to enteric coated mycophenolic acid and try to reduce the cumulative immunosuppressive dosage according to the

immunological risk. If the diarrhea still persists, patients underwent a colonoscopy to look for other causes such as CMV colitis or mycophenolate toxicity.

LHBT is a standard diagnostic test for LM in clinical practice. However, 2 potential limitations should be mentioned: Small intestine bacterial overgrowth (SIBO) with lactose fermentation and H₂ production in the small intestine could potentially lead to a false positive LHBT. However, SIBO and LM can be distinguished, since the resulting H₂ peak will be early in the former (small bowel peak), but delayed and more prominent in the latter (colonic peak).¹⁸ Furthermore, in a variable fraction of individuals (2-43%, <10% in most studies) the bowel flora does not produce H₂, leading to a false negative LHBT.¹⁶

Limitations of LTT include fluctuations of blood sugar levels for instance due to impaired glucose tolerance, diabetes or other influences.¹⁹ The small increase in blood sugar levels required for a positive test will thus result in an inferior sensitivity and specificity of LTT, therefore this test is not recommended as a routine diagnostic test for LM.^{10,20} However, since limitations of LHBT and LTT are largely nonoverlapping these tests can complement each other. In our study we found a perfect agreement of both tests regarding LM, arguing for the validity of our results.

Importantly, in the noro group but not in the control group, patients reported symptoms after lactose ingestion, suggesting that patients with chronic norovirus infection in fact suffer from LI. For most individuals with LM, a blinded challenge with 25 g of lactose does not result in any symptoms.²¹ Therefore, symptoms after lactose ingestion in individuals with LM are likely due to the concomitant presence of visceral hypersensitivity, for instance due to irritable bowel syndrome (IBS). This was also suggested in a blinded controlled study where a challenge with 20

g lactose resulted in typical symptoms in 47% of patients with IBS but only in 22% of control patients.²² Postinfectious IBS has been reported as a complication of viral gastroenteritis.²³ Our patients were not formally tested for the presence of IBS. However, our data suggest that visceral hypersensitivity can also complicate chronic norovirus infection.

Secondary LM has been reported in patients with intestinal inflammation due to other chronic inflammatory conditions including Crohn's disease.^{24,25} In a rat model of mucositis, lactose digestion was severely reduced, along with downregulation of lactase mRNA and protein levels, while glucose absorption remained intact.²⁶ In this study, other intestinal disaccharidases were also downregulated, suggesting that lactase might be a marker for a more general derangement of digestive enzymes. Clearly, an isolated downregulation of lactase does not explain chronic diarrhea in norovirus infected patients since most individuals with primary LM are free of symptoms. Therefore, further mechanistic studies regarding the expression of lactase levels and other disaccharidases are clearly needed.

We are conscious that our study has several limitations. First, the sample size is small which is due to the low prevalence of symptomatic chronic norovirus infection. Secondly, since our study focused on RTRs the results might not be extrapolated to other immunocompromised disease states in which chronic norovirus infection is a relevant concern.⁸ Thirdly, the investigators were not blinded for the results of norovirus PCR before lactose tolerance testing and no blinded placebo control was done as suggested by an NIH conference addressing LI.²⁷ Thus, placebo effects for the development of symptoms cannot be totally excluded. Finally, no additional tests regarding intestinal malabsorption were performed and our study does not provide a comprehensive information regarding all aspects of intestinal malfunction in norovirus infection.

Conclusion

This is the first study to show a positive association between chronic norovirus infection and secondary LM in RTRs. Future studies should address whether a lactose-reduced diet might be of therapeutic benefit.

ACCEPTED

References

1. Ahmed SM, Hall AJ, Robinson AE, et al. Global prevalence of norovirus in cases of gastroenteritis: a systematic review and meta-analysis. *Lancet Infect Dis.* 2014;14:725-730.
2. Robilotti E, Deresinski S, Pinsky BA. Norovirus. *Clin Microbiol Rev.* 2015;28:134-164.
3. Glass RI, Parashar UD, Estes MK. Norovirus gastroenteritis. *N Engl J Med.* 2009;361:1776-1785.
4. Westhoff TH, Vergoulidou M, Loddenkemper C, et al. Chronic norovirus infection in renal transplant recipients. *Nephrol Dial Transplant.* 2009;24:1051-1053.
5. Roos-Weil D, Ambert-Balay K, Lanternier F, et al. Impact of norovirus/sapovirus-related diarrhea in renal transplant recipients hospitalized for diarrhea. *Transplantation.* 2011;92:61-69.
6. Schorn R, Hohne M, Meerbach A, et al. Chronic norovirus infection after kidney transplantation: molecular evidence for immune-driven viral evolution. *Clin Infect Dis.* 2010;51:307-314.
7. Capizzi T, Makari-Judson G, Steingart R, Mertens WC. Chronic diarrhea associated with persistent norovirus excretion in patients with chronic lymphocytic leukemia: report of two cases. *BMC Infect Dis.* 2011;11:131.
8. Bok K, Green KY. Norovirus gastroenteritis in immunocompromised patients. *N Engl J Med.* 2012;367:2126-2132.
9. Coste JF, Vuiblet V, Moustapha B, et al. Microbiological diagnosis of severe diarrhea in kidney transplant recipients by use of multiplex PCR assays. *J Clin Microbiol.* 2013;51:1841-1849.

10. Misselwitz B, Pohl D, Fruhauf H, Fried M, Vavricka SR, Fox M. Lactose malabsorption and intolerance: pathogenesis, diagnosis and treatment. *United European Gastroenterol J*. 2013;1:151-159.
11. Itan Y, Jones BL, Ingram CJ, Swallow DM, Thomas MG. A worldwide correlation of lactase persistence phenotype and genotypes. *BMC Evol Biol*. 2010;10:36.
12. Enattah NS, Sahi T, Savilahti E, Terwilliger JD, Peltonen L, Jarvela I. Identification of a variant associated with adult-type hypolactasia. *Nat Genet*. 2002;30:233-237.
13. Gonzalez-Galan V, Sanchez-Fauquier A, Obando I, et al. High prevalence of community-acquired norovirus gastroenteritis among hospitalized children: a prospective study. *Clin Microbiol Infect*. 2011;17:1895-1899.
14. Rana SV, Bhasin DK, Vinayak VK. Lactose hydrogen breath test in Giardia lamblia-positive patients. *Dig Dis Sci*. 2005;50:259-261.
15. Khanna R, Vinayak VK, Mehta S, KumKum, Nain CK. Giardia lamblia infection in immunosuppressed animals causes severe alterations to brush border membrane enzymes. *Dig Dis Sci*. 1988;33:1147-1152.
16. Gasbarrini A, Corazza GR, Gasbarrini G, et al. Methodology and indications of H₂-breath testing in gastrointestinal diseases: the Rome Consensus Conference. *Aliment Pharmacol Ther*. 2009;29 Suppl 1:1-49.
17. Newcomer AD, McGill DB, Thomas PJ, Hofmann AF. Prospective comparison of indirect methods for detecting lactase deficiency. *N Engl J Med*. 1975;293:1232-1236.
18. Romagnuolo J, Schiller D, Bailey RJ. Using breath tests wisely in a gastroenterology practice: an evidence-based review of indications and pitfalls in interpretation. *Am J Gastroenterol*. 2002;97:1113-1126.

19. Arola H. Diagnosis of hypolactasia and lactose malabsorption. *Scand J Gastroenterol Suppl.* 1994;202:26-35.
20. Terjung B, Lammert F. [Lactose intolerance: new aspects of an old problem]. *Dtsch Med Wochenschr.* 2007;132:271-275.
21. Suarez FL, Savaiano DA, Levitt MD. A comparison of symptoms after the consumption of milk or lactose-hydrolyzed milk by people with self-reported severe lactose intolerance. *N Engl J Med.* 1995;333:1-4.
22. Yang J, Deng Y, Chu H, et al. Prevalence and presentation of lactose intolerance and effects on dairy product intake in healthy subjects and patients with irritable bowel syndrome. *Clin Gastroenterol Hepatol.* 2013;11:262-268 e261.
23. Zanini B, Ricci C, Bandera F, et al. Incidence of post-infectious irritable bowel syndrome and functional intestinal disorders following a water-borne viral gastroenteritis outbreak. *Am J Gastroenterol.* 2012;107:891-899.
24. Mishkin B, Yalovsky M, Mishkin S. Increased prevalence of lactose malabsorption in Crohn's disease patients at low risk for lactose malabsorption based on ethnic origin. *Am J Gastroenterol.* 1997;92:1148-1153.
25. Kirschner BS, DeFavaro MV, Jensen W. Lactose malabsorption in children and adolescents with inflammatory bowel disease. *Gastroenterology.* 1981;81:829-832.
26. Fijlstra M, Rings EH, Verkade HJ, van Dijk TH, Kamps WA, Tissing WJ. Lactose maldigestion during methotrexate-induced gastrointestinal mucositis in a rat model. *Am J Physiol Gastrointest Liver Physiol.* 2011;300:G283-291.

27. Suchy FJ, Brannon PM, Carpenter TO, et al. NIH consensus development conference statement: Lactose intolerance and health. *NIH Consens State Sci Statements*. 2010;27:1-27.

ACCEPTED

Figure 1. Study flow chart

LHBT, lactose H₂ breath test; LTT, lactose tolerance test

Figure 2. Lactose H₂ breath test

Exhaled H₂ content before and after ingestion of 25 g lactose

Figure 3. Lactose tolerance test

Serum glucose before and after ingestion of 25 g lactose

ACCEPTED

Figure 1

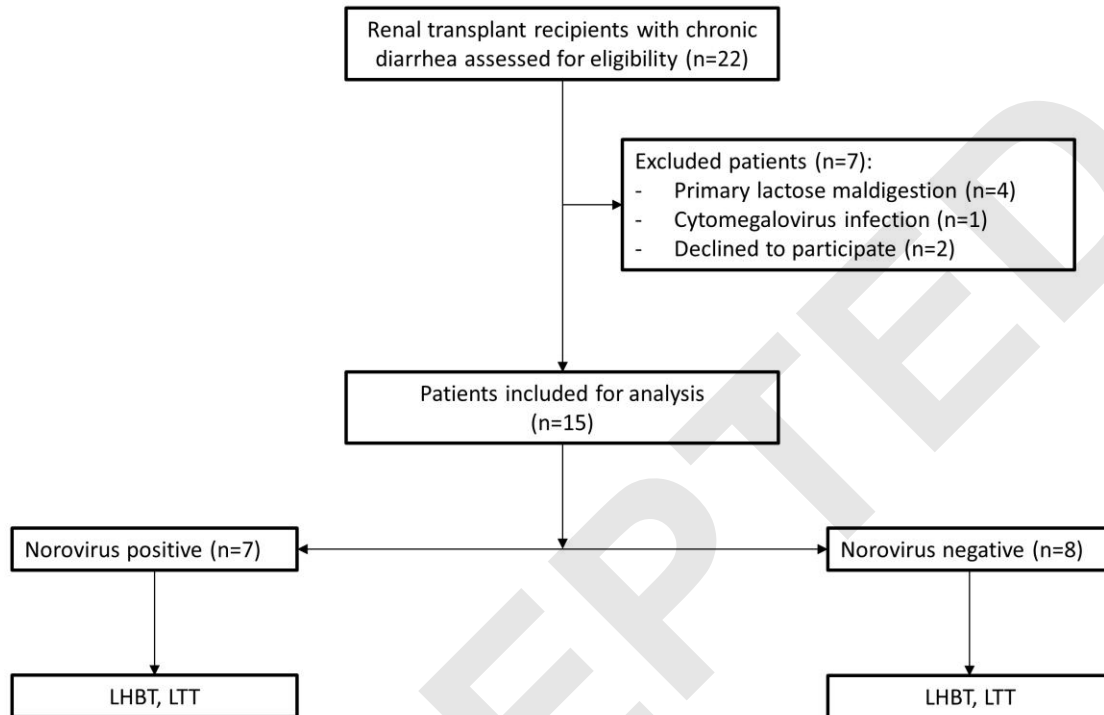


Figure 2

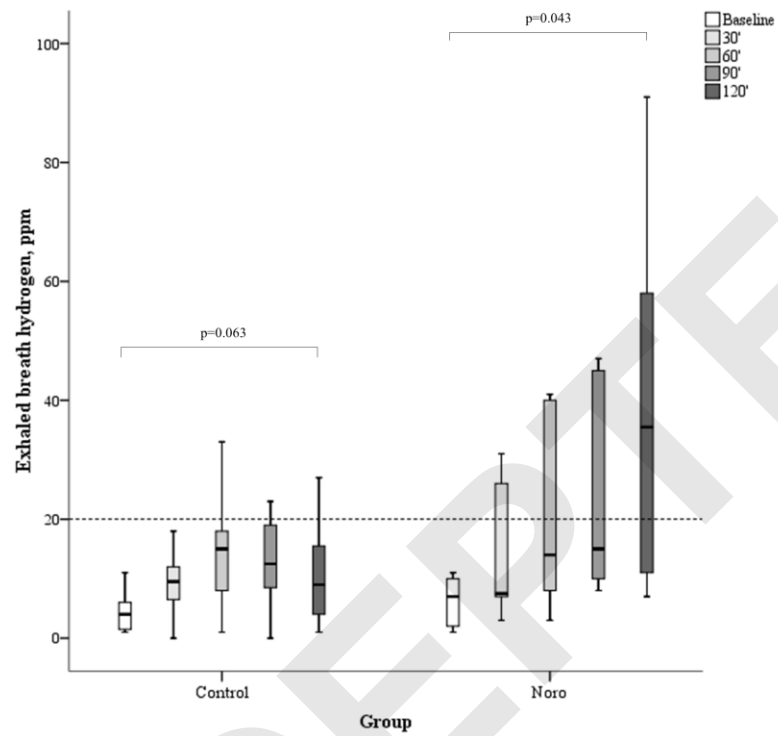


Figure 3

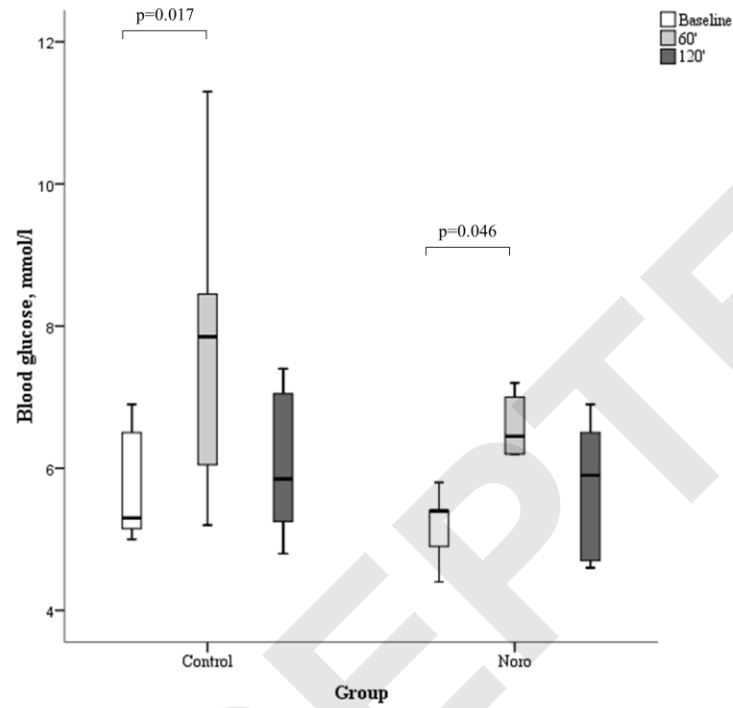


Table 1. Baseline characteristics

	Control group (n=8)	Noro group (n=7)	<i>p</i> - value
Age, years	52 (47, 60)	50 (34, 56)	0.46
Male sex	6 (75)	4 (57)	0.46
BMI, kg/m ²	24.8 (23.2, 26.5)	22.2 (17.0, 24.4)	0.09
Time since transplantation, years	2 (1-4)	6 (2-9)	0.23
RTx/RPTx	7/1	5/2	0.44
Immunosuppression, n (median daily dose; through level)			
- Prednisone	2 (5 mg)	1 (10 mg)	0.87
- Azathioprine	0	1 (150 mg)	0.69
- Mycophenolate	8 (1250 mg)	6 (1000 mg)	0.23
- Ciclosporine	1 (150 mg; 51 µg/l)	1 (90 mg; 50 µg/l)	0.78
- Tacrolimus	7 (4 mg; 7.3 µg/l)	6 (3.3 mg; 6.8 µg/l)	0.85
CMV risk constellation			
- High (D+/R-)	4 (50)	0 (0)	0.013
- Intermediate (D+/R+ or D-/R+)	2 (25)	7 (100)	
- Low (D-/R-)	2 (25)	0 (0)	
CMV PCR negative	8	7	0.46
Duration of diarrhea, weeks	13 (6-22)	35 (24-46)	0.038
Diabetes mellitus	3 (38)	2 (12)	0.31

Data are presented as median (interquartile range), or numbers (percent). BMI, body mass index; CMV, cytomegalovirus; PCR, polymerase chain reaction; RTx, renal transplantation; RPTx, combined renal-pancreatic transplantation

ACCEPTED

Table 2. Baseline laboratory values

	Control group	Noro group	<i>p</i>-
	(n=8)	(n=7)	value
Hemoglobin, g/l	135 (118, 147)	122 (106, 133)	0.46
Leucocytes, G/l	6.1 (4.8, 6.9)	4.1 (4.0, 6.0)	0.12
Lymphocytes, G/l	1.42 (0.93-1.61)	1.00 (0.57-1.05)	0.041
Alanine transaminase, U/l	20 (14, 24)	22 (17, 37)	0.28
Alkaline phosphatase, U/l	56 (55, 81)	80 (55, 92)	0.71
Estimated glomerular filtration rate*, ml/min/1.73 m ²	48 (43, 58)	43 (40, 48)	0.40
Potassium, mmol/l	4.1 (4.0, 4.1)	4.4 (3.8, 4.8)	0.40
Sodium, mmol/l	140 (138, 140)	141 (139, 143)	0.28
Glucose, mmol/l	5.4 (4.8, 6.0)	5.1 (5.0, 5.5)	0.69
C-reactive protein, mg/l	0.9 (0.6, 1.5)	0.4 (0.3, 0.5)	0.054

Data are presented as median (interquartile range). * CKD-EPI equation.

Table 3. Test results of LHBT and LTT

	Noro group (n=7)	Control group (n=8)
LHBT / LTT positive	7 (100)*	1 (12.5)*
GI symptoms after lactose	6 (85.7) [#]	0 (0) [#]

Values are displayed in n (%). * p=0.001; [#] p=0.057.

GI, gastrointestinal; LHBT, lactose H₂ breath test; LTT, lactose tolerance test

Verdankung

Herrn Dr. med. Daniel Franzen möchte ich für die freundliche Überlassung des hochinteressanten Themas herzlich danken. Ich verdanke ihm darüber hinaus jede erdenkliche, hilfreiche Unterstützung und viele anregende Diskussionen. Jede Phase dieser Arbeit wurde von ihm intensiv sowie professionell begleitet. Sein kompetenter Rat und seine Hilfe kamen mir in zahlreichen Angelegenheiten sehr zugute.

Mein besonderer Dank gilt auch Herrn Dr. med. Marco Bonani. Jederzeit gewährte er mir bei der Planung, Durchführung und Auswertung der vorliegenden Arbeit außerordentlich sachkundige, erfahrene und wertvolle Unterstützung.

Dem Herrn PD Dr. med. Benjamin Misselwitz sei herzlich gedankt für die hilfreichen Tipps und Ergänzungen zu den gastroenterologischen Beiträgen in dieser Arbeit.

Ein ganz besonderer Dank geht an die Mitarbeitenden Brigitta Gabathuler und Diana Jovanovic des Funktionslabors für die außerordentlich gute Zusammenarbeit.

Begleittext zur Publikation

Von Rajha Messias Fabrizio Pereira

Hintergrund/Fragestellung

Eine chronische Norovirus-Infektion mit Viruspersistenz geht oft mit einer chronischen Diarrhoe und Abdominalbeschwerden einher. Deren Pathogenese ist bislang nicht genau geklärt. Bei Infektionen durch Rotaviren und Giardia lamblia wurde eine Laktosemaldigestion bzw. eine Schädigung der Dünndarmschleimhaut mit einem Verlust von Disaccharidasen nachgewiesen, was zu entsprechenden Symptomen mit Diarrhoe, Blähungen und Abdominalschmerzen führt (Gonzalez-Galan, Sanchez-Fauquier et al., 2011; Khanna, Vinayak et al., 1988; Rana, Bhasin et al., 2005). Aufgrund von dieser Erkenntnis wollten wir die Hypothese überprüfen, ob der Diarrhoe bei einer chronischen Norovirus-Infektion ebenfalls ein Laktasemangel zugrunde liegen könnte.

Der primäre Endpunkt dieser Arbeit war es daher, die Inzidenz einer Laktosemaldigestion bei Patienten mit symptomatischer chronischer Norovirusinfektion im Vergleich zu Patienten mit chronischer Diarrhoe aufgrund einer anderen Ursache zu untersuchen. Da chronische Norovirusinfektionen v.a. bei immunsupprimierten Patienten auftreten, wurden ausschliesslich nierentransplantierte Patienten in die Studie eingeschlossen.

Patientenkollektiv, Methoden, Datenauswertung und Statistik

Patientenkollektiv

Im Zeitraum zwischen Juli 2013 und März 2015 wurden alle erwachsenen Nierentransplantationspatienten am Universitätsspital Zürich, bei denen eine chronische Norovirusinfektion mit Diarrhoe vorlag, angefragt, an der vorliegenden prospektiven, parallellaufenden Kohortenstudie teilzunehmen. In Übereinstimmung mit den Kriterien der Weltgesundheitsorganisation (WHO) wurde eine chronische Diarrhoe als das Vorliegen von mindestens drei Stuhlentleerungen am Tag über einen Zeitraum von mindestens vier Wochen definiert. Das Kriterium für eine chronische Norovirusinfektion war der Nachweis von Noroviren durch die Polymerasekettenreaktion (PCR) in mehr als zwei Stuhlproben über einen Zeitraum von mindestens einem Monat. Als Vergleichsgruppe dienten nierentransplantierte

Patienten mit einer chronischen Diarrhoe ohne Nachweis einer Norovirusinfektion oder einer anderen Infektion.

Ausschlusskriterien für die Teilnahme an der Studie waren:

- Eine Darminfektion durch Infektionserreger wie Viren (ausser Norovirus), Bakterien (Salmonellen, Campylobacter und Shigellen) oder Parasiten (besonders *Gardia lamblia*, *Microspora* spp., and *Cryptospora* spp.) nachgewiesen durch Polymerasekettenreaktion (PCR), Kultur oder mikroskopische Untersuchung. Eine Cytomegalievirusinfektion wurde mittels PCR im Blut sowie Koloskopie mit Histologie ausgeschlossen.
- Eine primäre adulte Laktosemalabsorption bzw. eine primäre Laktoseintoleranz definiert als molekulargenetischer Nachweis des CC-Genotyps des C/T-13910-Polymorphismus vor dem 5'-Ende des Laktasegens.
- Eine bekannte Galaktosämie oder Notwendigkeit einer galaktosearmen Diät.

Kriterien für die Aufnahme in die Kontrollgruppe waren:

- Alter > 18 Jahre
- Zustand nach einer Nierentransplantation
- Vorliegen einer chronischen Diarrhoe unklarer Ursache
- Ausschluss einer Norovirusinfektion sowie anderer viraler, bakterieller und parasitärer Darminfektionen durch PCR bzw. Kultur oder mikroskopische Untersuchung.

Der Auftrag des Doktorierenden beim Patientenkollektiv bestand darin sämtliche Patienten anhand der gewählten Kriterien in einer Excel-Tabelle festzuhalten und zu rekrutieren.

Methode

Laktose H₂-Atemtest und Laktosetoleranztest

Bei allen Patienten wurde ein Laktose H₂-Atemtest und ein oraler Laktosetoleranztest durchgeführt. Das Kriterium für die Diagnose einer Laktosemaldigestion war ein pathologisches Ergebnis beim H₂-Atemtest und/oder dem oralen Laktosetoleranztest.

Laktose H₂-Atemtest

Der Laktose H₂-Atemtest (LH₂BT) wurde gemäß Newcomer et al. (Newcomer, McGill et al., 1975) durchgeführt. Nach nächtlichem Fasten über mindestens 12 Stunden wurde eine basale Atemprobe (Nüchternprobe) abgenommen. Während der Fastenzeit durften die Patienten Wasser trinken. Die übliche Medikation wurde während der gesamten Untersuchung weitergeführt. Nach der Abnahme der Nüchternprobe tranken die Patienten 50 g Laktose gelöst in 250 ml Wasser. Proben von 20ml der endexpiratorischen Atemluft wurden jeweils 30, 60, 90 und 120 Minuten nach der oralen Laktosebelastung mit dem AlveoSampler™ Lactose Kit (Quintron Instrument Co., Milwaukee, WI, USA) gesammelt. Eine H₂-Konzentration von mehr als 20 parts per million (ppm) wurde als Kriterium für das Vorliegen einer Laktosemaldigestion gewertet (Gasbarrini, Corazza et al., 2009; Newcomer, McGill et al., 1975). Während der Untersuchung durften die Patienten nicht essen, sonst aber ihren üblichen Aktivitäten nachgehen ohne den Laborraum zu verlassen. Das Trinken von Wasser war jederzeit erlaubt. Die Untersuchung wurde in einem gut gelüfteten Raum durchgeführt. Zigarettenrauch, frisch gestrichene Wände oder sonstige frisch gestrichene Gegenstände und organische Lösungsmittel waren nicht vorhanden. Die Proben der Atemluft wurden in speziell hierfür konstruierten Taschen gesammelt, die zusammen mit dem Messgerät von der Klinik für Gastroenterologie USZ zur Verfügung gestellt wurden. Die Wasserstoff-Konzentration wurde dann mit dem Gerät 12i Microlyser (Quintron Instrument Co., Milwaukee, WI, USA) gemessen. Während der Untersuchungsperiode wurde aufgezeichnet, wie oft es zu Stuhlentleerungen und zum Abgang von Darmgasen kam, oder ob der Patient an Abdominalbeschwerden litt.

Laktosetoleranztest

Gleichzeitig an die oben beschriebene orale Verabreichung wurden die Blutzuckerwerte im Kapillarblut bei 0, 60 und 120 Minuten mit Hilfe eines Blutzuckermessgeräts (Accu-Chek Roche Diabetes Care (Schweiz) AG) bestimmt. Ein Anstieg des Blutzuckerspiegels von weniger als 1,1 mmol/l in Verbindung mit dem Auftreten von Abdominalbeschwerden wurde als beweisend für das Vorliegen einer Laktoseintoleranz definiert (Newcomer, McGill et al., 1975).

Der Auftrag des Doktorierenden bestand zunächst in der Motivation und Einplanung der Patienten im Gastroenterologie-Labor am USZ sowie die Patienten am Untersuchungstag am Empfang abzuholen und mit ihnen die oben beschriebenen Untersuchungen im Labor selbstständig durchzuführen, nachdem die Einverständniserklärung eingeholt worden war. Die entstandenen Daten wurden in einem zuvor selber erstellten, für diese Arbeit zweckmässigen Protokoll erfasst. Alle Patienten wurden durch den Doktorierenden während der ganzen Untersuchung persönlich betreut. Im Falle eines positiven Testergebnisses wurde in Absprache mit einem Oberarzt der Nephrologie eine Empfehlung zur Ernährungsberatung dem Patienten unterbreitet und gegebenenfalls Instruktionen über die genetische Untersuchung erteilt sowie einholen der Unterschrift dazu veranlasst.

Datenauswertung und Statistik

Bei der deskriptiven Darstellung der Basisdaten wurde bei metrischen Parametern Median und Interquartilenbereich (IQR) berechnet. Die Berechnung von Unterschieden zwischen beiden Studiengruppen wurde bei metrischen Daten ein zweiseitiger Mann-Whitney U-Test eingesetzt. Bei kategorialen Variablen wurde die absolute Häufigkeit (n) und die relative Häufigkeit (%) bestimmt. Zur Überprüfung der Unabhängigkeit kategorialer Variablen wurde der Chi-Quadrat-Test eingesetzt.

Die Wasserstoffkonzentration in der Ausatemungsluft und der Blutzuckerspiegel wurden als Median und Interquartilenbereich (IQR) angegeben. Unterschiede zwischen verschiedenen Zeitpunkten wurden mit Hilfe des Wilcoxon-Rangsummentests für verbundene Stichproben berechnet. Als Signifikanzniveau wurde bei allen Test $p < 0,05$ festgelegt. Alle statistischen Berechnungen und grafischen Darstellungen wurden unter Verwendung des Programms SPSS Statistics für Windows, Version 22 (IBM Corporation, Armonk, NY) erstellt.

Der Auftrag des Doktorierenden war das selbstständige Erstellen der deskriptiven Statistik sowie einen Flow-Chart über den Patientenfluss zu erstellen. Zudem wurde die induktive Statistik mit Hilfe von Dr. med. Franzen durchgeführt.

Resultate

Von den etwa achthundert Patienten in der Kohorte der erwachsenen Nierentransplantationspatienten des Universitätsspitals Zürich wurden insgesamt 15 Patienten in die Studie eingeschlossen. Sieben dieser Patienten hatten eine chronische symptomatische Norovirus-Infektion mit Diarrhöe (Noro-Gruppe). Die Kontrollgruppe umfasste acht Patienten. Die Zusammensetzung des Patientenkollektivs ist in der Publikation Figure 1 (Study flow chart) dargestellt.

Die demografischen und klinischen Basisdaten sind in Table 1 zusammengefasst. „Baseline characteristics“, die laborchemischen Basisdaten sind in Table 2 ersichtlich. Die Zytomegalie-Hochrisiko-Konstellation war in der Kontrollgruppe signifikant häufiger als in der Noro-Gruppe ($p = 0,013$). Bei allen anderen Variablen (Alter, Geschlecht, Art und Dosierung der Immunsuppression, Häufigkeit eines Diabetes mellitus, Laborwerte) gab es keinen signifikanten Unterschied zwischen der Noro-Gruppe und der Kontrollgruppe. Während des Untersuchungszeitraums erhielt kein Patient Antibiotika.

In der Noro-Gruppe fiel der Laktose H₂-Atemtest und der Laktosetoleranztest bei allen Patienten positiv aus, wobei die H₂-Konzentration in der Ausatemungsluft 120 Minuten nach der Laktosegabe bei über 20 ppm lag. In der Kontrollgruppe fielen bei lediglich einem Patienten sowohl der Laktose H₂-Atemtest als auch der Laktosetoleranztest positiv aus. Bei den anderen sieben Patienten der Kontrollgruppe ergaben der Laktose H₂-Atemtest und der Laktosetoleranztest Normalwerte.

Bei sechs der sieben Patienten in der Noro-Gruppe (85,7%) traten nach der Laktosegabe abdominelle Beschwerden auf. Hierbei handelte es sich bei drei Patienten um eine Diarrhoe und bei drei Patienten um Meteorismus. In der Kontrollgruppe traten bei keinem Patienten Abdominalbeschwerden auf. Hierzu zählte auch der Patient mit einem positiven Laktose H₂-Atemtest und Laktosetoleranztest.

Aufgrund des Laktose H₂-Atemtest und Laktosetoleranztest (Tabelle 1) war die Laktosemaldigestion in der Noro-Gruppe signifikant häufiger als in der Kontrollgruppe (Odds Ratio 75%, 95% CI 2.6, 2153, $p = 0.01$).

Tabelle 1 Testergebnisse des Laktose H₂-Atemtests und Laktosetoleranztests (vgl. Publikation)

	Noro-Gruppe (n=7)	Kontroll-Gruppe (n=8)
LHBT / LTT positive	7 (100)	1 (12.5)
LHBT / LTT negative	0 (0)	7 (87.5)

Konklusion

Die mögliche Korrelation zwischen Norovirusinfektion und Laktasemangel spricht dafür, dass die Diarrhoe bei immunsupprimierten Patienten mit einer chronischen Norovirusinfektion durch eine sekundäre Laktosemaldigestion hervorgerufen oder unterhalten werden könnte. Zur abschließenden Klärung dieses Zusammenhangs ist eine placebokontrollierte Doppelblindstudie an einer größeren Patientenzahl erforderlich. Hierbei sollte auch die Auswirkung einer laktosearmen Diät untersucht werden.

Der Auftrag des Doktorierenden war schlussendlich das Erstellen eines ersten Drafts des Manuskriptes.

Curriculum Vitae

Rajha Messias Fabrizio Pereira

18.10.1976	Geboren in Zürich, Schweiz
04/83 – 07/89	Primarschule, Zürich
07/89 – 07/92	Sekundarschule, Zürich
07/92 – 01/98	Kantonsschule Rämibühl, MNG, Zürich
11/96 – 06/01	Night-Auditor Claridge Hotel Tiefenau, Zürich
07/97 – 08/97	Disponent Häfeli Haustechnik AG, Schönenwerd
02/98 – 05/98	Rekrutenschule, Fribourg
06/98 – 07/98	Unteroffiziersschule, Fribourg
10/98 – 10/10	Medizinstudium, dipl. Arzt, Universität Zürich
11/99 – 12/99	Assistent PSP Swiss Property, Glattbrugg
12/00 – 07/01	Assistent SG Rüeegg Bank, Zürich
06/05 – 02/06	Assistent Credit Suisse (CS, CSFB, CSAM), Zürich
04/07 – 12/12	Dozent Dickerhof AG, Emmenbrücke
11/09 – 01/11	Dozent SFK AG, Zürich
12/10 – 01/13	Dozent Sanita GmbH, Derendingen
02/11 – 12/12	Assistent Dumo AG, Spreitenbach
02/11 – 04/13	Dozent Bénédict AG, Zürich
02/12 – 12/15	Dozent Juventus-Woodtli Stiftung, Zürich
09/12 – 02/13	Didaktikkurs 1 (Modul 1), Zürich
10/12 – 05/13	SVEB 1, Zürich
06/13 – 08/16	Bereichsleitung medizinische Grundlagen, Bodyfeet AG, Thun
01/16 – 07/16	CAS Unternehmensführung UZH, Zürich
seit 01/13	Publikation im Rahmen der Dissertation USZ, UZH, Zürich
seit 01/2014	Vorstandsmitglied OdA AM, Zürich
seit 01/2013	Dozent Bodyfeet AG, Rapperswil
seit 11/2012	Berufsschullehrer ZAG, Winterthur